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## Parasites of fishes from Laurel Creek, Ontario

KALMAN MOLNAR,\* GEORGE HANEK† AND C. H. FERNANDO

*Department of Biology, University of Waterloo, Waterloo, Ontario N2L 3G1, Canada*

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Two-hundred and nine fish of 13 species from Laurel Creek, Ontario, were examined for parasites between May and October 1973. Eighty-four species of parasites (22 of Protozoa, 24 of Monogenea, 17 of Digenea, 11 of Cestoda, 4 of Nematoda, 1 of Hirudinea, 1 of glochidia, and 4 of Crustacea) were collected and are listed and discussed.

### I. INTRODUCTION

Intensive parasitological investigations on fishes have been made in Ontario. Prominent among these are the studies by Bangham (1941), Bangham & Venard (1946), Dechtiar (1966), Freeman (1964), and Mizelle & Donahue (1944) from Algonquin Park, by Bangham (1955) and Mavor (1916) from Lake Huron, by Bangham & Hunter (1939) and Hunter & Bangham (1932, 1933) from Lake Erie, by Hanek & Fernando (1971a,b, 1972a,b, 1973) and Tedla & Fernando (1969a,b,c,d) from Lake Ontario, and by Dechtiar (1972) from the Lake of the Woods. Smaller habitats, particularly the streams, have been neglected. Laurel Creek, conveniently located within the immediate vicinity of the University of Waterloo, was therefore selected to fill a gap. The present study is an extension of the unpublished work of Kakonge (1970, 1972) on the ecology of fishes and other parasitological aspects of the stream.

### II. MATERIALS AND METHODS

Laurel Creek is a small, typical southern Ontario stream which flows into the Grand River, Lake Erie watershed. It is composed of pools alternating with shorter riffle sections and is 2.5-4.0 m wide and 50-120 cm deep at normal flow. The substrate of the pools is mud and plant debris while elsewhere the substrate is composed of fine sand or a mixture of sand and small gravel, except for the riffles which are composed of larger gravel and rubble. During the study period (May-October, 1973), the water temperature was warmer than 15°C, with a peak range of 24-28°C in July and August. The pH, dissolved oxygen concentration and conductivity ranged from 7.5-8.8, 40-190% saturation and 375-545 µmhos at 20°C respectively. Two-hundred and nine fish of 13 species (Table I) were subjected to a complete parasitological examination during the study period. They were caught with a small hand seine and by traps, and were brought to the laboratory alive and examined immediately. Conventional parasitological techniques were employed in the dissection of all fish and in preservation and preparation of the parasites. Hosts were identified using the keys and systematics of Hubbs & Lagler (1964), while systematic assignment of species follows the arrangement in Special Publication No. 6 of the American Fisheries Society (1970). Specimens of all the parasites found have been deposited in the collection of one of us (G.H.).

\* On leave from Veterinary Medical Research Institute, Academy of Sciences, Budapest, Hungary.

† Present address: Ministry of Agriculture and Fisheries, Nassau, P.N., Bahamas.

TABLE I. Protozoan parasites collected from the Laurel Creek fishes

Parasites	Locations	Hosts									
		<i>Bodotrius concava</i> Davis, 1947					<i>Spiromyces</i> sp.				
<i>Eimeria degustii</i> Molinar & Fernando, 1974	Spleen	20	15	10	15	15	15	15	10	22	20
<i>Eimeria freemani</i> Molinar & Fernando, 1974	Kidney	—	—	x	—	—	—	—	—	—	—
<i>Eimeria hybognathi</i> Molinar & Fernando, 1974	Intestine	11	—	—	—	—	—	—	—	—	—
<i>Eimeria ictaluri</i> Molinar & Fernando, 1974	Intestine	—	—	—	—	—	—	—	—	—	—
<i>Eimeria iroquoina</i> Molinar & Fernando, 1974	Intestine	6	1	—	3	—	—	—	—	—	—
<i>Eimeria laureliensis</i> Molinar & Fernando, 1974	Intestine	—	x	x	—	x	—	xx	—	—	—
<i>Noropis cornutus</i> (Mitchill)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Platyphales promelas</i> (Rahneseque)	Habits	5	—	—	—	—	—	—	—	—	—
<i>Platyphales notatus</i> (Rahneseque)	Habits	—	—	x	—	—	—	—	—	—	—
<i>Noctomitus bimaculatus</i> (Kirtland)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Catostomus commersoni</i> (Lacepede)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Ictalurus nebulosus</i> (Lesuer)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Micropodus salmoides</i> (Lacepede)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Ambloplites rupestris</i> (Rahneseque)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Etheostoma exile</i> (Girard)	Gills	—	—	—	—	—	—	—	—	—	—
<i>Perca flavescens</i> (Mitchill)	Gills	—	—	—	—	—	—	—	—	—	—

<i>Eimeria</i> spp.	Intestine		6	2	6
<i>Sphaerosporu</i> sp.	Kidney, urinary ducts	1 xxx	— —	— xx	— xx
<i>Chloromyxum catostomi</i> Kudo, 1920	Gallbladder	4 xx	— xx	— xx	— xx
<i>Myxidium</i> sp.	Gallbladder	3 xxx	1 —	1 xxx	1 xxx
<i>Myxobolus hyborhynchi</i> Fatham <i>et al.</i> , 1939	Kidney	3 xxx	2 xx	2 —	2 —
<i>Myxobolus transversalis</i> Fatham <i>et al.</i> , 1939	Kidney	1 xx	5 x	— x	— x
<i>Myxobolus</i> sp.	Gills	6 xx	2 —	2 —	2 —
<i>Myxobolus</i> spp.	Gallbladder, intestine	— xx	— xx	— xx	— xx
<i>Hemigryya exilis</i> Kudo, 1929	Gills	— x	3 —	2 —	1 —
<i>Glugea</i> sp.	Intestine, gallbladder	1 xxx	1 —	— xx	2 —
<i>Ichthyophthirius multifiliis</i> Fouquet, 1876	Fins	— xx	— x	— xx	— xx
<i>Aplosooma</i> spp.	Gills, fins	— xx	1 —	1 xx	2 —
<i>Trichodina</i> spp.	Gills, fins	7 x	6 x	5 x	11 x
<i>Trichodina</i> sp.	Urinary ducts	1 xxx	2 —	x x	10 —

Numerator = number of fish infested, denominator = infestation: x, slight; xx, moderate; xxx, high.

TABLE II. Monogenea collected from the Laurel Creek fishes

Parasites	Locations	No. examined						
			20	15	10	15	15	15
<i>Dactylogyrus attenuatus</i> Mizelle & Klucka, 1953	Gills	7						
<i>Dactylogyrus bychowskyi</i> Mizelle, 1937	Gills							
<i>Dactylogyrus banghami</i> Mizelle & Donahue, 1944	Gills	5						
<i>Dactylogyrus bulbis</i> Mueller, 1938	Gills							
<i>Dactylogyrus cornutus</i> Mueller, 1938	Gills							
<i>Dactylogyrus dubius</i> Mizelle & Klucka, 1953	Gills							
<i>Dactylogyrus</i> spp.	Gills							
<i>Actinocleidus fusiformis</i> (Mueller, 1934)	Gills							
<i>Cleidodiscus aleutus</i> Mueller, 1938	Gills							

(Lacépède)  
Micropterus salmoides  
(Rafinesque)  
Ambloplites rupestris  
(Lacépède)  
Micropterus salmoides  
(Lesauv.)  
Ictalurus nebulosus  
(Girard)  
Ethelostoma exile  
(Mitchilli)  
Percula flavaeens

a

<i>Cleidodiscus pricei</i> Mueller, 1936	Gills	17	<u>1-50(21)</u>	<u>2</u>	<u>1-2</u>	<u>1-24(7)</u>
<i>Cleidodiscus senior</i> Mueller, 1937	Gills					
<i>Urocleidus adpectus</i> Mueller, 1936	Gills					
<i>Urocleidus chautauquensis</i> (Mueller, 1938)	Gills					
<i>Urocleidus dispar</i> (Mueller, 1936)	Gills					
<i>Urocleidus furcatus</i> (Mueller, 1937)	Gills					
<i>Urocleidus principalis</i> (Mizelle, 1936)	Gills					
<i>Urocleidus</i> sp.	Gills					
<i>Otomacrum lanceatum</i> Mueller, 1936	Gills	3	<u>4-20(11)</u>	<u>1</u>	<u>1</u>	<u>1-5</u>
<i>Gyrodactylus eltheostoma</i> Wellborn & Rogers, 1967	Fins					
<i>Gyrodactylus hoffmani</i> Wellborn & Rogers, 1967	Fins	3	<u>1-5(3)</u>	<u>2</u>	<u>2</u>	<u>1-5</u>
<i>Gyrodactylus macrochiri</i> Hofman & Putz, 1964	Fins					
<i>Gyrodactylus nebulosus</i> Kritsky & Mizelle, 1968	Fins					
<i>Gyrodactylus stunkardi</i> Kritsky & Mizelle, 1968	Fins					
<i>Gyrodactylus</i> spp.	Fins, gills	9	<u>1-14(4)</u>	<u>9</u>	<u>9</u>	<u>1-30(10) 1-7(3)</u>

Numerator = number of fish infested, denominator = range followed by average of infestation in parenthesis.

TABLE III. Other parasites (Digenea, Cestoda, Nematoda, Hirudinea, Crustacea, glochidia) collected from the Laurel Creek fishes

Parasites	Locations	Hosts										
		No. examined										
		20	15	10	15	15	15	10	22	20	22	20
<i>Flagiporus sinisini</i> Mueller, 1934	Gallbladder	2		2				1				3
<i>Triganodistomum attenuatum</i> Mueller & Van Cleave, 1932	Intestine		1-9									
<i>Phyllobothrium lysteri</i> Miller, 1940	Urinary ducts							4				
<i>Azygia angusticauda</i> (Stafford, 1904)	Stomach											
<i>Glossidium geminum</i> (Mueller, 1930)	Intestine											
<i>Allocreadium lobatum</i> Wallin, 1909	Intestine											
* <i>Rhipidocotyle</i> sp.	Fins											
* <i>Diplostomum spathaceum</i> (Rud., 1819)	Lenses	11	14	6	3		2			3		3
* <i>Diplostomum scheuringi</i> Hughes, 1929	Vitreous chamber	1-15(5)	1-5(3)	1-6(4)	1-3(2)	3-5(4)	4-5	5	3	1-7(3)		
* <i>Tetracotyle</i> spp.	Brain, eyes, abdominal cavity						2	1	1	5		
* <i>Ornithodorostomum pychochelis</i> (Faust, 1917)	Brain, viscera	20	13	10*		15	1-1	7	7	1-3(2)	1-1	1-4(2)
		50-250 (130)	4-50(29) (60)	30-100 (55)	30-100 (55)	13-50 (31)	1-20(10) (55)	18-100 (55)				

* <i>Poecilodiplostomum minimum</i>							
<i>centrarchi</i> (MacCallum, 1921) Hoffman, 1958	Mesenteries	20 20-50 (38)	9 10-30 3-40(15) (15)	15 10-30 1-8(3)	4 8-100 (40)	15 8-100 (40)	1 1-20(10) 1-30(17)
* <i>Poecilodiplostomum minimum minimum</i> (MacCallum, 1921) Hoffman, 1958	Fins, skin, gills	10 1-50(17)	5 5 12 9	14 14	5 9	3 3-25(10) (32) (3)	14 1-20(5) 1-80(16)
* <i>Uvulifer ambloplitis</i> (Hughes, 1927)	Fins, skin	3 4-20(11)				3 1-3(2)	14 1-20(5) 1-80(16)
* <i>Nexacanthus pyriformis</i> Chandler, 1951	Skin, musculature					6 6-50(20) (3)	
* <i>Clinostomum marginatum</i> (Rind., 1819)	Skin					1 1-3(2) (2)	
* <i>Ribeiroia ondatrae</i> (Price, 1931)						4 1-3(2) (2)	
<i>Biacetabulum biloculoides</i>	Intestine						
Mackiewicz & McCrae, 1965							
<i>Glaridacris longae</i> (Lamont, 1921)	Intestine						
<i>Hinterella nodulosa</i>	Intestine						
Mackiewicz & McCrae, 1962							
<i>Monobothrium hunteri</i>	Intestine						
Mackiewicz, 1963							
<i>Caryophyllidea</i> sp.	Intestine						
* <i>Ligula intestinalis</i> (L., 1758)	Abdominal cavity						
<i>Schistocerca solidus</i>	Abdominal cavity						
(Müller, 1776)							
<i>Corallobothis fimbriatum</i>	Intestine						
Essex, 1927							
Corallobothis sp.	Intestinal serosa						
* <i>Cysticercus dilepis</i>	Gallbladder						
<i>unilateralis</i> (Rud., 1819)							
* <i>Cysticercus dilepis</i> sp.	Gallbladder						
<i>Philometra cylindracea</i>	Abdominal cavity, swimmbadder						
Ward & Magath, 1916							
						4 1-3(2)	

TABLE III—continued

Parasites	Locations	Hosts									
		20	15	10	15	15	15	10	22	20	10
<i>Philotrema</i> sp.	Abdominal cavity, swimmbbladder							7			
* <i>Spiroxiis contortus</i> (Rud., 1819)	Intestinal serosa							1-5(4)	1		1
* <i>Elastrongylides</i> sp.	Abdominal cavity								—	7	1
<i>Actinobedella triangulata</i> Moore, 1924	Gills										
<i>Glochidium</i> spp.	Fins, gills	9	2	3	1	7	6	1	1	2	4
		1-16(3)	1-5	3-7(5)	2	3-8(5)	1-26(7)	1	—	2	2-8(3)
<i>Argulus catostomi</i> Dana & Herrick, 1837	Fins										
<i>Ergasilus caeruleus</i> Wilson, 1911	Gills										
<i>Ergasilus centrarchidarum</i> Wright, 1882	Gills										
<i>Ergasilus cyprinaceus</i> Rogers, 1969	Gills	4	1	—	—	1	—	1	5	2	
		1-2	1						1-20(10)	2-2	

Numerator = number of fish infested, denominator = range followed by average of infestation in parenthesis.  
\*, larval stage.

### III. RESULTS AND DISCUSSION

Eighty-four species of parasites (22 of Protozoa, 24 of Monogenea, 17 of Digenea, 11 of Cestoda, 4 of Nematoda, 1 of Hirudinea, 1 of glochidia, and 4 of Crustacea) (Tables I, II, III) were obtained, and all the species of parasites reported in this study constitute new distribution records. Twelve new host records are as follows: *Bodomonas concava* and *Henneguya exilis* for *Ictalurus nebulosus*; *Eimeria iroquoina* for *Nocomis biguttatus*, *Notropis heterolepis*, *Pimephales notatus* and *P. promelas*; *Chloromyxum catostomi* for *Notropis cornutus* and *Semotilus atromaculatus*; *Myxobolus transversalis* for *Hybognathus hankinsoni* and *Notropis heterolepis*; *Cysticercus dilepis unilateralis* for *Micropterus salmoides*; and *Schistocephalus solidus* for *Perca*

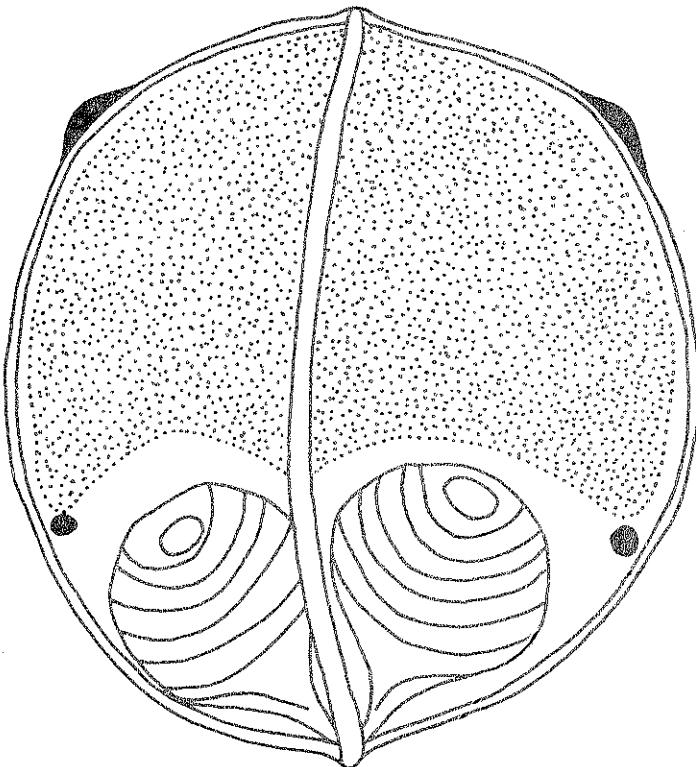


FIG. 1. *Sphaerospora* sp.

*flavescens*. Furthermore, *Cysticercus dilepis unilateralis* is reported for the first time from North America, and several species of the genera *Dactylogyrus* and *Gyrodactylus* (listed in Table II as *Dactylogyrus* spp. and *Gyrodactylus* spp.) are new for science and their detailed description is now in preparation (Hanek, Molnar, & Fernando, in press). When examining the intestine of various fish species we have often noticed flagellates with eight flagella resembling *Hexamita salmonis* (Moore, 1923), which is a common parasite of North American salmonids (Davis, 1953). Kulda & Lom (1964) have done much work on this group of parasites which they consider to belong to the genus *Spironucleus* Lavier, 1936. Fantham *et al.* (1939) described *Sphaerospora notropis* from the oral epithelium and muscles of *Notropis cornutus*. The spores of *Sphaerospora* sp. noticed during the present study (Fig. 1) were found in the ureter and

resemble the species commonly found in the kidneys of European and Asian fishes. The short description is as follows: spores spherical with somewhat protrusive anterior pole and sutural ridge; valves smooth. Small lateral outgrow present on the slightly flattened posterior pole. Polar capsules slightly pyriform. Length of spores 9·0–10·4  $\mu$ , width 9·0–9·6  $\mu$ , length of capsules 3·0–3·3  $\mu$ , diameter 2·5–2·7  $\mu$ . The parasitofauna of the fishes of Laurel Creek was characterized by the high intensity of infestation with metacercaria, those of *Ornithodiplostomum ptychocheilus* and *Posthodiplostomum minimum* being the most frequent parasites. The high intensity of infestation can be explained by the rich fauna of birds in this region. Furthermore, the metacercaria of *Ornithodiplostomum ptychocheilus* were encountered in about equal numbers in the abdominal and cranial cavities; the only exception being *Notropis heterolepis* in which the metacercaria infested the cranial cavity only. Also, it was found that the metacercaria of *P. minimum* were localized on the serosas in the abdominal cavities of cyprinids, while this parasite was found infesting only the livers of the centrarchids and *Perca flavescens*. This finding supports the opinion of

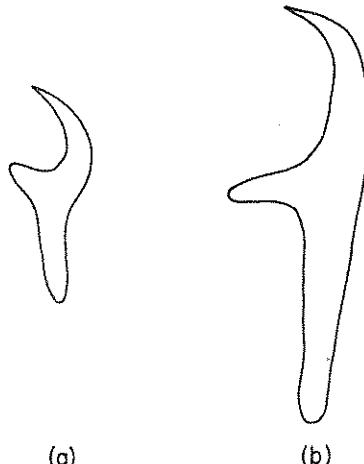


FIG. 2. *Cysticercus dilepidis* sp. (a) lower row hook, (b) upper row hook.

Hoffman (1958) that there are two subspecies of *P. minimum*: *P. m. minimum* infesting the cyprinids and *P. m. centrarchi*, infesting the centrarchids. Infestation with *Diplostomum spathaceum* was significant in *Ictalurus nebulosus*, and like Ali & Hanyu (1964) we have found this parasite infesting the small hernias of the lenses, which they called 'multiple lenses'. The infestation with *Neascus* of *Uvulifer ambloplites* was also fairly high. In some cases we have found these metacercaria together with *Neascus pyriformis*; the later species differs from *U. ambloplites* in having a pyriform inner cyst wall, and according to Chandler (1951) it may be identical with the metacercaria of *Uvulifer semicircumcisus*. Two morphologically different types of *Cysticercus* were collected from the gallbladder of *Micropterus salmoides*. One of them was identical with the larval form of *Dilepis unilateralis* and its short description is as follows: body oval, 500–520  $\mu$  long and 220–250  $\mu$  wide. Head with four simple suckers and single terminal trunk armed with two rows of hooks; 10 hooks in each row. Length of upper row hooks 23–24  $\mu$ , lower row hooks 10–11  $\mu$ . The short description of the other type, *Cysticercus dilepidis* sp. follows: body oval, 520–560  $\mu$

long and 220–250  $\mu$  wide. Head with four simple suckers and single terminal trunk armed with two rows of hooks; also 10 hooks in each row. Length of upper row hooks [Fig. 2(b)] 37–39  $\mu$ , lower row hooks [Fig. 2(a)] 19–21  $\mu$ . Despite the frequent occurrence of glochidia, we did not deal with this group in detail, because these parasites were intensively studied by Kakonge (1972) who investigated the ecology and pathology of *Anodonta plana*, *Anodontoides ferussacianus*, *Lasmigona compressa* and *Lampsilis* sp.

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